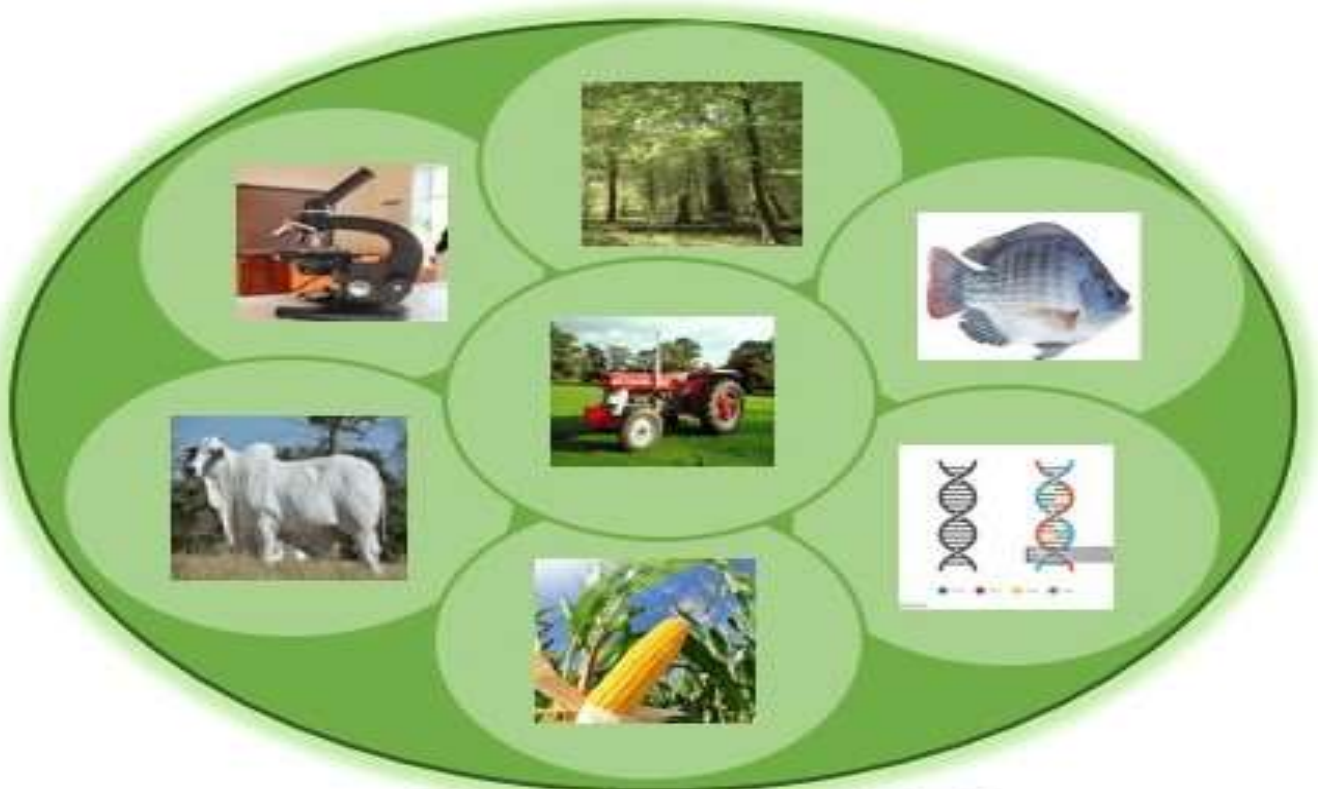




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PERFORMANCE OF HYBRID AFRICAN CATFISH (*Heteroclarias*) FINGERLINGS FED DIETS CONTAINING *Aphodius rufipes* LARVAE MEAL AND *Hermetia illucens* LARVAE MEALS

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ABSTRACT

This study investigated the growth performance and health of hybrid African catfish (*Heteroclarias*) fingerlings fed diets containing varying inclusion levels (0 %, 15 %, 30 %, and 45 %) of *Aphodius rufipes* larvae meal (ARLM) and *Hermetia illucens* larvae meal (HILM). Seven (7) iso-nitrogenous diets (40 % crude protein) were formulated and ten (10) fingerlings were stocked per aquarium (45 × 30 × 24 cm³) for a 70-day feeding trial. Growth parameters (weight gain, specific growth rate, feed conversion ratio), haematological indices (packed cell volume, haemoglobin, platelet count), proximate and mineral compositions of fish carcasses were assessed. One-way analysis of variance (ANOVA) and Duncan's Multiple Range Test (DMRT) were used for statistical analysis ($p < 0.05$). Results showed no significant differences ($p > 0.05$) in weight gain, average daily gain, protein efficiency ratio or feed conversion ratio among treatments. However, the 30 % ARLM diet yielded the highest weight gain (25.26 g). Significant differences ($p < 0.05$) were observed in protein retention, apparent net protein utilization, protein retention efficiency, survival rate, haematological indices and carcass compositions (proximate and minerals). The study revealed that ARLM and HILM could be incorporated into *Heteroclarias* fingerlings diets up to 45 % without negatively impacting growth or health.

Keywords: Performance, *Heteroclarias*, Diets, Fingerlings

Introduction

Aquaculture is the growing of aquatic organisms in confined areas (Huntington *et al.*, 2017) which play an important role in economies worldwide, in both developed and developing countries (Subasinghe *et al.*, 2009). Aquatic food consumption globally is projected to reach 21.4 kg per capita in 2031, up from the baseline of 20.5 kg per capita (Mair *et al.*, 2023). Per capita, consumption will increase in all continents except Africa, the

region with the fastest-growing population (FAO, 2022). By 2030, projections indicate a significant and substantial increase in aquaculture production compared to capture fisheries, with an estimated 6 million tonnes (Mair *et al.*, 2023). However, if growth in aquaculture can be sustained, it is likely to meet the increasing demand for aquatic food supplies by supplying more than 50 percent of the total aquatic food consumption (Subasinghe *et al.*, 2009; Tran *et al.*, 2022).

Fish farming is a branch of aquaculture that grows fish in an enclosed system (Walia and Kaur, 2023). Fish is highly nutritious, rich in micronutrients, minerals, essential fatty acids and protein (Selamoglu and Naeem, 2023). Fish contributes to or exceeds 50 percent of total animal protein intake in some small Islands and other developing States like Nigeria (Norman *et al.*, 2019). The observed expansion of fish farming directly correlates with the progressive intensification of its production systems (Azim and Little, 2007). Intensive production systems in this sector rely heavily on manufactured feeds designed to meet the specific nutritional requirements of the cultured fish species (Gabriel *et al.*, 2007). Fish feed constitutes up to 70 % of the variable operating costs in commercial fish farming. The cost of fishmeal, a crucial protein source in fish feed, is a major determinant of the overall feed production expenses (Jannathulla *et al.*, 2019). Conventional fish feeds predominantly utilize both plant and animal-derived (fishmeal, soybean meal and maize meal) protein sources (Jimoh *et al.*, 2014). However, economic viability necessitates the development and utilization of cost-effective feed formulations (non-conventional materials) for sustainable fish farming practices (Shipton *et al.*, 2013).

Aphodius rufipes larvae, an insect from Scarabaeidae, are found within cattle dung in natural savannah and grassland grazing ecosystems (Bake *et al.*, 2021). Analysis reveals that edible *Aphodius rufipes* larvae feeding on cattle dung possess high protein levels and other essential nutrients (Alamu *et al.*, 2013; Tang *et al.*, 2019). The larvae of *Hermetia illucens* are saprophagous, consuming a wide variety of decaying organic matter, including animal manure and human excreta, with minimal substrate disruption (Suriya *et al.*, 2024). It produces a biomass with approximately 42 % protein and up to 35 % fat content (Stamer *et al.*, 2014). The high protein

content and favorable amino acid profile of *Hermetia illucens* larvae suggest their potential as a novel animal feed ingredient (Cammack and Tomberlin, 2017). *Hermetia illucens* larvae demonstrate significant potential as a sustainable protein source due to its efficient bioconversion of low-value organic substrates into nutritionally rich biomass (Danieli *et al.*, 2019). The readily available larvae, possessing a high-protein content and favourable amino acid profile, represent a promising candidate species for animal feed applications (Cammack and Tomberlin, 2017). Therefore, this study was conducted to evaluate the growth responses of hybrid African catfish (*Heteroclarias*) fingerlings fed diets containing *Aphodius rufipes* larvae meal (ARLM) and *Hermetia illucens* larvae meal (HILM).

Materials and Methods

Study Area

This study was conducted at the Department of Water Resources, Aquaculture and Fisheries Technology (WAFT), Old Teaching and Research Farm, Bosso Campus, Federal University of Technology (FUT) Minna, Niger State, Nigeria. Minna exhibits a sub-humid climate characterized by a mean annual rainfall of 1284 mm and a distinct dry season lasting from November to March (Ojanuga, 2006). Mean maximum temperatures remain consistently high, averaging approximately 32°C, with peak values observed in March and June. Geographically, Minna is situated (9°31'N, 6°30'E) within Nigeria's Southern Guinea savanna vegetation belt of Nigeria (Lawal *et al.*, 2012).

Experimental Ingredients

Fishmeal (FM), soybean meal (SBM), maize meal (MM), starch, vegetable oil and a vitamin/mineral premix were obtained from Kure Ultra-modern Market, Bosso Local Government Area, Niger State. The soybean was slightly toasted. The larvae of *Aphodius*

rufipes (dung beetles) were collected from compacted cattle dung in Wuyagi village (9°8'31"N, 5°51'36"E), Lavun Local Government Area, Niger State, Nigeria. Following dissection to remove unwanted materials, the larvae were euthanized and cleaned by immersion in 60°C water for 30 minutes. Subsequently, samples were oven-dried at 100°C for 8 hours. *Hermetia illucens* (black soldier fly) eggs were sourced from a reputable farm in Minna, Nigeria and incubated in an open container with moistened maize

chaff (*Dusa*). After 48 hours, hatched larvae were fed a diet of fruit waste (watermelon, pineapple and cucumber) for three weeks. Larvae were harvested using a water-based flotation technique, sieved (0.5 mm mesh size) and oven-dried at 100°C for four hours. All ingredients were individually processed, milled, sieved and packed in an airtight polyethene bag until needed. The ingredients were analysed for proximate and mineral compositions (Table 1).

Table 1
Proximate and Mineral Compositions of the Major Feed Ingredients

Ingredients/ Parameters	FM	ARLM	HILM	SBM	MM	
						Proximate Compositions (%)
Crude protein	56.88	39.50	28.00	43.01	9.62	
Ether extract	12.98	14.83	15.76	21.39	6.71	
Crude fibre	0.00	14.65	7.70	6.22	2.55	
Ash content	13.37	11.55	23.62	5.36	1.54	
Moisture content	10.43	9.29	4.41	3.73	9.05	
Nitrogen free extract	6.34	10.18	20.51	20.29	70.53	
					Mineral Compositions (g/kg)	
Sodium	1.52	1.68	1.78	0.06	0.09	
Potassium	8.56	7.10	7.26	1.76	0.83	
Phosphorus	0.21	0.13	0.19	0.09	0.06	
Calcium	1.88	1.14	1.66	0.21	1.26	
Magnesium	0.77	0.39	0.76	0.51	0.43	

FM = Fishmeal, ARLM = *Aphodius rufipes* larvae meal, HILM = *Hermetia illucens* larvae meal, SBM = Soybean meal, MM = Maize meal

Formulation of the Experimental Diets

Seven (7) iso-nitrogenous diets of 40 % crude protein (CP) were formulated (Table 2) for *Heteroclaris* fingerlings using linear programming (Microsoft Excel) with varying inclusion levels of insects meal and designated as T1 (0 %: Control), T2 (15 % ARLM), T3 (30 % ARLM), T4 (45 % ARLM), T5 (15 % HILM), T6 (30 % HILM) and T7 (45 %

HILM). Feed ingredients were independently weighed using a weighing (Golden Mettler 2000L: Model) balance. A moist dough was created by homogenizing the ingredients in a plastic bowl with added steam for gelatinization. This dough was then pelletized (2 mm diameter) using a manual pelletizer and oven-dried at 80°C for 40 minutes. The formulated diets were then analysed for proximate and mineral compositions (Table 3).

Table 2. Compositions of the Experimental Diets

Treatments/Ingredients	T1	T2	T3	T4	T5	T6	T7
<i>Aphodius rufipes</i> larvae meal	0.00	15.00	30.00	45.00	0.00	0.00	0.00
<i>Hermetia illucens</i> larvae meal	0.00	0.00	0.00	0.00	15.00	30.00	45.00
Fishmeal	64.76	54.35	43.93	33.51	52.55	40.34	28.13
Maize meal	10.00	10.00	10.00	10.00	10.00	10.00	10.00
Soybean meal	5.00	5.00	5.00	5.00	5.00	5.00	5.00
Vegetable oil	2.92	2.02	1.12	0.21	2.37	1.81	1.26
Vitamin/Mineral premix	5.00	5.00	5.00	5.00	5.00	5.00	5.00
Cellulose	12.32	8.63	4.95	1.28	10.08	7.85	5.61
Total	100.00	100.00	100.00	100.00	100.00	100.00	100.00
Energy kcal/100 g	271.76	271.81	271.84	271.77	271.80	268.73	271.81

Table 3. Proximate and Mineral Compositions of the Experimental Diets

Treatments/Parameters	T1	T2	T3	T4	T5	T6	T7
Proximate Compositions (%)							
Crude protein	39.21	39.56	39.44	38.96	39.61	39.06	39.15
Ether extract	10.97	13.84	10.21	9.13	13.96	10.51	11.12
Crude fibre	2.52	2.66	3.52	3.96	3.02	4.02	3.78
Ash content	11.06	13.56	14.12	13.33	14.12	15.75	15.42
Moisture content	6.88	5.18	5.56	3.61	4.43	5.73	4.08
Nitrogen free extract	29.36	25.20	27.15	31.01	24.86	24.93	26.45
Mineral Compositions (g/kg)							
Sodium	0.48	0.45	0.53	0.79	0.43	0.86	0.69
Potassium	1.02	0.96	0.86	1.12	1.43	1.10	1.33
Phosphorus	0.11	0.14	0.12	0.11	0.15	0.20	0.19
Calcium	0.29	0.21	0.22	0.25	0.29	0.26	0.25
Magnesium	0.52	0.57	0.49	0.62	0.76	0.54	0.43

Experimental Fish

The hybrid African catfish (*Heteroclaris*) fingerlings were obtained from a reputable fish farm in New Bussa, Niger State and transported in an open plastic container to the experimental site. They were acclimatized upon arrival in an outdoor nursery tank (150

× 150 × 75 cm³) for two weeks before the experiment commenced. They were fed a conditioning diet (vital fish feed) twice daily.

Determination of Growth Parameters

The growth parameters were calculated using the formulae:

Weight gain (WG) = Final weight (Wf) – Initial weight (Wi) of the fish.

$$\text{Specific growth rate (SGR)} = \frac{\ln W_f - \ln W_i}{T} \times 100$$

Where: ln Wf = logarithm of the final weight, ln Wi = logarithm of the initial weight and T = experimental duration.

Feed conversion ratio (FCR) = $\frac{\text{Weight of feed fed (dry weight)}}{\text{Weight gain of fish (wet weight)}}$.

Average daily gain (ADG) = $\frac{\text{Mean final body weight} - \text{Mean initial body weight}}{\text{Number of days of the experiment}}$.

Survival rate (SR) = $\frac{N_f}{N_i} \times 100$

Where: N_f = number of alive at the end of the experiment, N_i = number stocked at the beginning of the experiment.

Protein retention (PR) = $\frac{\text{Protein gain}}{\text{Protein fed}} \times 100$

Protein efficiency ratio (PER) = $\frac{\text{Weight gain}}{\text{Protein fed}} \times 100$

Apparent net protein utilization (ANPU) = $\frac{\text{Carcass protein gain (g)}}{\text{Protein fed (g)}} \times 100$

Protein retention efficiency (PRE) = $\frac{\text{Protein retained}}{\text{Protein consumed}}$

Haematological Indices

The fish blood samples (Table 4) were collected from each treatment group by bleeding the fish from the caudal fin using a dissecting blade at the beginning and end of the feeding trial as described by Vijayan and Moon (1992); and Stoskopf (1993). The collected blood samples were immediately placed into plastic tubes containing ethylene diamine tetra-acetic acid (EDTA) as an anticoagulant and transported to the Veterinary Hospital laboratory (Niger State Ministry of Livestock and Fisheries Minna) for analysis. The mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH) and mean corpuscular haemoglobin concentration (MCHC) were calculated using the formulae described in Doig and Zhang (2017).

MCV (fl) = $PCV/RBC \times 10$; MCH (pg) = $Hb/RBC \times 10$ and $MCHC$ (g/dl) = $Hb/PCV \times 100$.

Biochemical Analysis

The samples of feed ingredients formulated diets and fish carcasses (Table 5) were taken to the WAFT laboratory for analysis of proximate and mineral compositions. The proximate analysis (crude protein, ether extract, moisture, ash and fibre content) was conducted according to AOAC (2000) methods. Nitrogen-free extract (NFE) was calculated by difference: $100 - (\text{crude protein} + \text{ether extract} + \text{moisture} + \text{ash} + \text{fibre})$. For mineral analysis, 2 g samples were weighed (Atom-110C balance) in a crucible and ash (M110 muffle furnace at 500°C for 4 hours), followed by acid digestion (10 ml HCl).

Table 4: Haematological Indices of *Heteroclarias* Fingerlings Fed the Experimental Diets for 70 Days

Treatments/ Parameters	PCV (%)	Hb (g/dl)	RBC ($\times 10^9/l$)	WBC ($\times 10^9/l$)	PLC (%)	MCV (fl)	MCH (pg)	MCHC (%)
Initial	13.67	4.56	2.28	0.28	102.67	60.03	20.01	33.34
T1	17.67 ^{bc}	5.89 ^{bc}	2.95 ^{bc}	0.95 ^{bc}	116.33 ^{ab}	59.96 ^a	19.98 ^a	33.32 ^a
T2	20.00 ^b	6.67 ^b	3.33 ^b	1.33 ^b	124.00 ^{ab}	60.06 ^a	20.03 ^a	33.35 ^a
T3	17.00 ^{bcd}	5.67 ^{bcd}	2.83 ^{bcd}	0.83 ^{bcd}	121.00 ^{ab}	60.00 ^a	20.00 ^a	33.33 ^a
T4	13.33 ^d	4.44 ^d	2.22 ^d	0.22 ^d	107.00 ^b	59.97 ^a	19.98 ^a	33.33 ^a
T5	15.00 ^{cd}	5.00 ^{cd}	2.50 ^{cd}	0.50 ^{cd}	106.00 ^b	60.00 ^a	20.00 ^a	33.33 ^a
T6	27.00 ^a	9.00 ^a	4.50 ^a	2.50 ^a	160.67 ^a	60.00 ^a	20.00 ^a	33.33 ^a
T7	21.00 ^b	7.00 ^b	3.50 ^b	1.50 ^b	131.00 ^{ab}	60.00 ^a	20.00 ^a	33.33 ^a
±SE	1.02	0.34	0.17	0.17	5.79	0.01	0.01	0.00

Values in the same column with different superscripts are significantly different ($p < 0.05$) from each other.

PCV = Packed cell volume, Hb = Haemoglobin, RBC = Red blood cell, WBC = White blood cell, PLC = Platelet count, MCV = mean corpuscular volume, MCH = Mean corpuscular haemoglobin, MCHC = Mean corpuscular haemoglobin concentration

Table 5: Proximate and Mineral Compositions of the Experimental Fish (Carcass: Dry Basis) Fed the Diets for 70 Days

Treatments/ Parameters	Proximate Compositions (%)									
	Initial	T1	T2	T3	T4	T5	T6	T7	±SE	
Crude protein	48.18	58.52 ^{ab}	54.83 ^{bcd}	60.74 ^a	56.74 ^{bc}	53.38 ^{cd}	51.46 ^d	58.25 ^{ab}	0.77	
Ether extract	9.16	10.37 ^d	11.03 ^{cd}	11.95 ^{bc}	12.29 ^{bc}	12.53 ^b	15.18 ^a	11.77 ^{bc}	0.34	
Ash content	18.18	16.91 ^{ab}	17.73 ^a	13.81 ^{bc}	13.78 ^{bc}	11.65 ^c	17.06 ^{ab}	14.40 ^{abc}	0.58	
Crude fibre	7.25	5.72 ^{de}	7.03 ^b	5.32 ^e	6.26 ^{bcd}	6.12 ^{cd}	6.58 ^{bc}	8.39 ^a	0.22	
Moisture content	6.15	5.28 ^b	5.50 ^{ab}	4.96 ^b	5.08 ^b	5.25 ^b	6.25 ^a	5.57 ^{ab}	0.12	
Nitrogen free extract	11.09	3.20 ^b	3.87 ^b	3.23 ^b	5.85 ^b	11.07 ^a	3.47 ^b	1.62 ^b	0.77	
		Mineral Compositions (g/kg)								
Sodium	0.44	0.48 ^c	0.51 ^b	0.55 ^{ab}	0.52 ^b	0.48 ^c	0.56 ^a	0.53 ^{ab}	0.01	
Potassium	1.06	1.28 ^{bc}	1.42 ^b	1.79 ^a	1.70 ^a	1.20 ^c	1.40 ^b	1.35 ^{bc}	0.05	
Phosphorus	0.11	0.11 ^b	0.11 ^{ab}	0.12 ^{ab}	0.12 ^a	0.09 ^c	0.12 ^{ab}	0.11 ^b	0.00	
Calcium	0.42	0.39 ^b	0.35 ^b	0.28 ^c	0.40 ^b	0.29 ^c	0.51 ^a	0.52 ^a	0.02	
Magnesium	0.2	0.40 ^a	0.34 ^a	0.21 ^b	0.34 ^a	0.23 ^b	0.36 ^a	0.21 ^b	0.02	

Values in the same row with different superscripts are significantly different ($p < 0.05$) from each other.

The resulting solution was filtered (50 ml volumetric flask) and analysed for sodium (Na) and potassium (K) using flame photometry (Jenway FF-200), phosphorus (P) using spectrophotometry (Jenway 741501), calcium

Experimental Conditions

The experiment was conducted using plastic aquaria ($45 \times 30 \times 24 \text{ cm}^3$). Following initial weighing to determine the mean weight (2.28 – 2.29 g), ten (10) fingerlings were stocked per aquarium at the commencement of the trial. The fingerlings were fed to satiation three times daily, with uneaten feed and faecal matter siphoned from aquaria each morning using a 7.5 mm diameter rubber hose. Sampling was done every two weeks. The fish were removed from each aquarium using a hand net and weighed using a sensitive weighing balance (Model: 2000L Golden Mettler). The water samples were collected bi-weekly and transported to the WAFT Department laboratory for analysis (Table 7) of dissolved oxygen (DO), temperature (T), pH, biological oxygen demand (BOD), alkalinity (ALK), water hardness (WH) and electrical conductivity (EC).

Data Analysis

The collected data were analysed using analysis of variance (ANOVA) with the aid of a statistical package for the social sciences (SPSS) version 27. Treatment means were compared using Duncan's Multiple Range Test (DMRT). Statistical significance was set at 5 % ($p < 0.05$).

Results and Discussions

The results in Table 1 showed that the *Aphodius rufipes* larvae meal (ARLM) contained a CP of 39.50 % and ether extract (EE) of 14.83 %. This was similar (34.41 % CP and 12.15 % EE) to that reported by Gana *et al.* (2022). The *Hermetia illucens* larvae meal (HILM) contains 28.00 % and 15.76 % CP and

(Ca) via EDTA titration (10 % KOH, casein indicator), and magnesium (Mg) via EDTA titration (ammonia buffer, Eriochrome black T indicator).

EE respectively. However, Wardhana *et al.* (2016); Loho and Lo (2023) report that the CP content of *Hermetia illucens* larvae typically falls within the range of 30 – 50 %, while the crude lipid content ranges from 29 – 32 %. These variations may be attributed to seasons, size, geographical locations and processing methods. The results in Table 6 show the growth performance of *Heteroclaris* fingerlings fed varying levels of ARLM and HILM based diets for 70 days. There was no significant difference ($p > 0.05$) in the initial mean weight, final weight and weight gain of the fish fed the experimental diets. Although the fish fed T3 (30 % ARLM) had the highest weight gain (25.26 g) while the T1 (0 %: Control) had the least weight gain of 17.02 g. This does not corroborate with the findings of Maranga *et al.* (2022) and Mundida *et al.* (2023). These researchers reported a significant difference ($p < 0.05$) among the treatments in *Clarias gariepinus* fingerlings when fed inclusion levels of HILM based diets. The observed differences could be because of different culture mediums, species, diet compositions and feeding periods. Weight gain in fish fingerlings is crucial for their survival and growth, ultimately impacting fish farming productivity and sustainability. No significant differences ($p > 0.05$) were observed among the treatment means in FCR. Mitra (2021) and Oteri *et al.* (2021) demonstrated that high-quality feeds directly improve FCR, a key factor in enhancing the economic viability of fish farming by minimizing feed costs and maximizing profitability. However, significant variations ($p < 0.05$) were observed among the treatment means in PR, ANPU and PRE. The percentage survival in all the fish fed dietary treatments ranged between 73.33 – 90.00 %

and they differ significantly. However, this was lower compared to the values (98.52 – 98.86 %) reported by Bake *et al.* (2021) while investigating the growth performance, body composition and apparent nutrient digestibility of hybrid catfish fingerlings fed with blended insect meal. The enhanced survival in the experimental group was likely due to the high palatability and nutritional quality of the

experimental diets, promoting improved health status and reduced mortality. High survival rates are essential for the economic sustainability of fish farming operations (Simasiku *et al.*, 2024). Figure 1 represents the growth curve of *Heteroclaris* fingerlings fed varying levels of ARLM and HILM based diets.

Table 6
 Growth Indices of *Heteroclaris* Fingerlings Fed the Experimental Diets for 70 Days

Treatments/ Parameters	T1	T2	T3	T4	T5	T6	T7	±SE
IW (g)	2.29	2.28	2.29	2.28	2.28	2.29	2.28	0.00
FW (g)	19.31	21.17	27.55	22.36	24.34	27.15	20.95	1.06
WG (g)	17.02 ^a	18.89 ^a	25.26 ^a	20.08 ^a	22.06 ^a	24.86 ^a	18.67 ^a	1.06
SGR (%)	3.04 ^a	3.17 ^a	3.54 ^a	3.26 ^a	3.30 ^a	3.53 ^a	3.16 ^a	0.06
FCR	1.96 ^a	1.74 ^a	1.31 ^a	1.60 ^a	1.67 ^a	1.31 ^a	1.69 ^a	0.08
PI	13.27 ^a	12.94 ^{ab}	12.86 ^{ab}	12.77 ^{ab}	12.81 ^{ab}	12.96 ^{ab}	12.44 ^b	0.07
ADG (g)	0.24 ^a	0.27 ^a	0.36 ^a	0.29 ^a	0.32 ^a	0.35 ^a	0.27 ^a	0.02
SR (%)	83.33 ^{ab}	80.00 ^{ab}	73.33 ^b	90.00 ^a	76.67 ^{ab}	86.67 ^{ab}	76.67 ^{ab}	1.81
PR	33.17 ^a	32.35 ^{ab}	32.16 ^{ab}	31.93 ^{ab}	32.03 ^{ab}	32.39 ^{ab}	31.10 ^b	0.19
PER	0.42 ^a	0.47 ^a	0.63 ^a	0.50 ^a	0.55 ^a	0.62 ^a	0.47 ^a	0.03
ANPU	21.45 ^{ab}	13.85 ^{cd}	26.07 ^a	17.81 ^{bc}	10.72 ^{de}	6.84 ^e	20.92 ^{ab}	1.53
PRE	4.41 ^{ab}	4.24 ^{bc}	4.73 ^a	4.45 ^{ab}	4.17 ^{bc}	3.97 ^c	4.68 ^a	0.07

Values in the same row with different superscripts are significantly different ($p < 0.05$) from each other

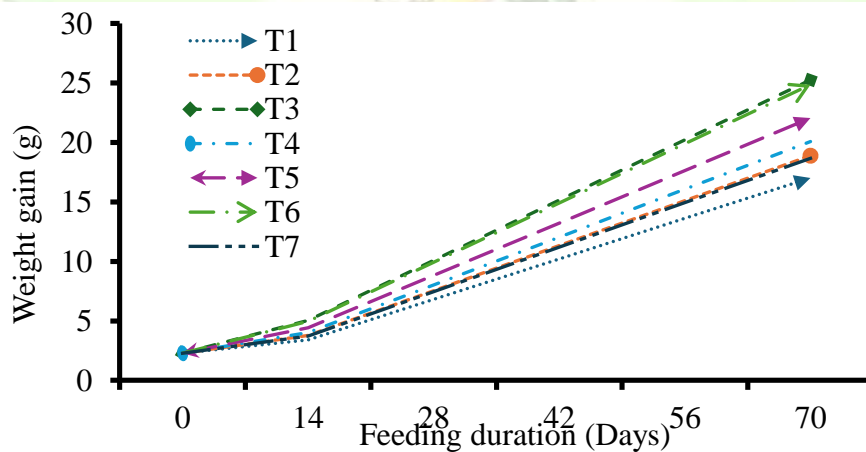


Figure 1: Growth Response Curve of *Heteroclaris* Fingerlings Fed Insects Meals for 70 Days

Haematological indices in African catfish fingerlings serve as valuable biomarkers reflecting physiological status and stress responses (Dahunsi and Oranusi, 2013; Muiyiwa *et al.*, 2020). The result (Table 4) revealed that the PCV, Hb and PLC differ significantly ($p < 0.05$) across the treatment means. The values recorded in this study are similar to those reported by Okore *et al.* (2016) while investigating the growth and haematological studies of African catfish (*Clarias gariepinus*) juveniles fed with housefly larva (*Musca domestica*) as a feed supplement. Fagbenro (2003) stated that poor water quality (high ammonia, low oxygen), temperature fluctuations and handling stress significantly alter haematological profiles. The Hb values ranged between 4.44 – 9.00 g/dl. Isaac *et al.* (2013) reported that haemoglobin transports oxygen to tissues for oxidative metabolism and carbon dioxide away from tissues. The PLC values recorded (106.00 – 160.67 %) were higher compared to the value (102.67 %) recorded at the initial (before the commencement of the experiment). Low platelet concentration suggests that the process of clot-formation (blood clotting) will be prolonged resulting in excessive loss of blood in the case of injury (Etim *et al.*, 2014).

The proximate and mineral compositions (Table 5) results revealed that the protein content (carcass) ranged between 51.46 – 60.74 %, 10.37 – 15.18 % for ether extract, 4.96 – 6.25 % for moisture content and differed significantly ($p < 0.05$) across the treatment means. This finding corroborates with the report of Muin *et al.* (2016) and Mundida *et al.* (2023), as they observed significant differences across the treatment means in fish body (carcass) compositions when fed inclusion levels of insect meals. The variations in body composition could indicate differences in fish species, age, diet and environmental conditions. Arbex *et al.* (2015) reported that

high protein content is essential for human health, while lipid content influences energy density and the presence of essential fatty acids. Hossain *et al.* (1992) reported that the ability to digest fat appears to be influenced by temperature and the level of fat in the diet. Therefore, the levels of these components are vital for assessing the quality and suitability of fish for human consumption or use in aquaculture feeds. The mineral compositions (carcass: dry basis) range from 0.48 – 0.56, 1.20 – 1.79, 0.09 – 0.12, 0.28 – 0.52 and 0.21 – 0.40 g/kg for sodium, potassium, phosphorus, calcium and magnesium respectively which differs significantly ($p < 0.05$) across the treatment means. Boyd and Pillai (1985); and Martínez-Valverde *et al.* (2000) stated that fish mineral composition varies according to species, age, diet and ambient water mineral content. Minerals are essential structural components of the skeletal system, constituents of organic compounds, enzyme activators and regulators of acid-base and osmotic balance (Soetan *et al.*, 2010). Selamoglu and Naeem (2023) reported that fish is a significant dietary source of essential minerals crucial for bone health, oxygen transport, immune function, thyroid hormone production and antioxidant defence.

Water quality is a critical determinant of fish farming success, significantly impacting fish growth, survival rates and overall health status (Simasiku *et al.*, 2024). The results (Table 7) showed no significant variations in water parameters (temperature, alkalinity and electrical conductivity) among the treatments fed the experimental diets. However, significant differences were observed across the treatment means in dissolved oxygen, biological oxygen demand, pH and water hardness. The experimental temperature range (25.90 – 27.50 °C) fell within the optimal range (24 – 30 °C), as reported by Santhosh and Singh (2007). The recorded dissolved oxygen

levels (5.53 – 6.77 mg/l) in this study were within the recommended range (4.98 – 6.70 mg/l) for maintaining fish health and promoting efficient feed conversion, as

reported by Mulugeta *et al.* (2024). Low levels of oxygen could lead to hypoxia, causing stress, reduced feed intake and impaired growth (Simasiku *et al.*, 2024).

Table 7: Water Quality Parameters (Monitored) of the *Heteroclaris* Fingerlings Fed the Experimental Diets for 70 Days

Treatments/ Parameters	T (°C)	pH	DO (mg/l)	BOD (mg/l)	ALK (mg/l)	WH (mg/l)	EC (µS/cm)
Initial	26.90	7.27	5.67	3.07	68.67	103.33	501.00
T1	26.80 ^a	6.88 ^b	6.30 ^{ab}	4.53 ^{ab}	75.33 ^a	113.67 ^{ab}	558.00 ^a
T2	26.20 ^a	6.81 ^b	6.77 ^a	4.97 ^a	73.33 ^a	111.67 ^b	540.00 ^a
T3	26.70 ^a	6.84 ^b	6.37 ^{ab}	4.57 ^{ab}	74.67 ^a	115.67 ^{ab}	552.00 ^a
T4	26.90 ^a	8.93 ^a	5.83 ^{ab}	4.10 ^b	71.33 ^a	115.33 ^{ab}	522.00 ^a
T5	25.90 ^a	6.80 ^b	5.53 ^b	3.83 ^b	78.67 ^a	121.00 ^{ab}	588.00 ^a
T6	27.50 ^a	6.90 ^b	5.87 ^{ab}	3.93 ^b	74.00 ^a	118.33 ^{ab}	546.00 ^a
T7	27.40 ^a	6.79 ^b	6.47 ^{ab}	4.07 ^b	79.33 ^a	128.67 ^a	594.00 ^a
±SE	0.12	0.21	0.14	0.11	1.11	1.94	9.97

Values in the same column with different superscripts are significantly different ($p < 0.05$) from each other.

T = Temperature, pH = Hydrogen ion concentration, DO = Dissolved oxygen, BOD = Biological oxygen demand, ALK = Alkalinity, WH = Water hardness EC = Electrical conductivity

Conclusion

This study demonstrated that incorporating *Aphodius rufipes* larvae meal (ARLM) and *Hermetia illucens* larvae meal (HILM) into *Heteroclaris* fingerling diets, at varying inclusion levels, did not significantly ($p > 0.05$) affect weight gain or feed conversion ratio. However, T3 (30 % ARLM) showed the highest weight gain (25.26 g) and T1 (0 %: control) the lowest (17.02 g). Therefore, these insect meals could be a suitable feed ingredient in *Heteroclaris* fingerling diet and could be included up to 45 %. Furthermore, it showed no negative impacts on the health and well-being of *Heteroclaris* fingerlings, as indicated by growth parameters and haematological indices, even at the highest inclusion level (45 %) of ARLM and HILM in the diets. The

haematological values could serve as baseline information for comparison in conditions of nutrient deficiency, physiology and health status of fish fingerlings. The variations observed in body proximate composition and mineral content highlight the need for careful consideration of insect meal incorporation to ensure optimal fish nutritional value.

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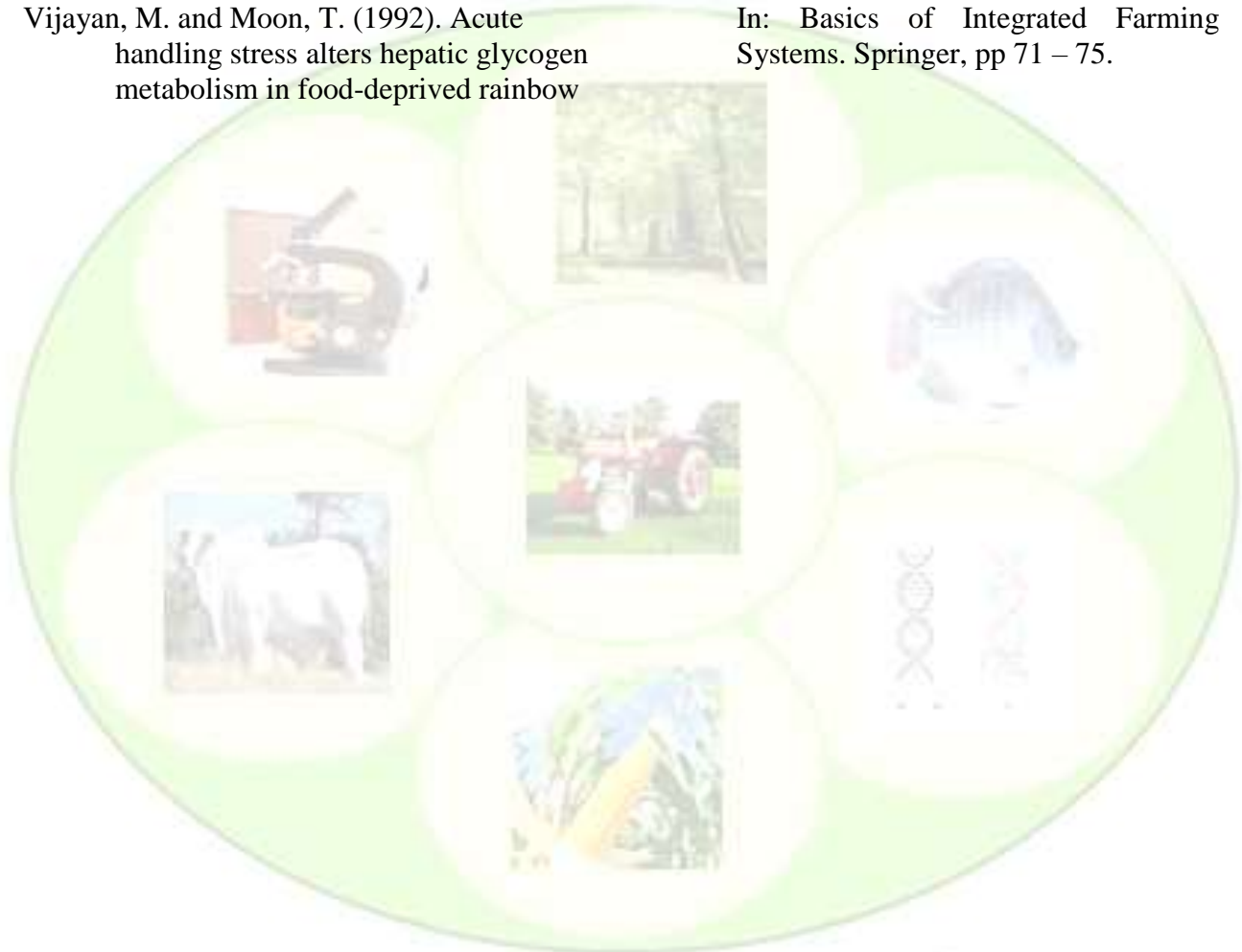
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